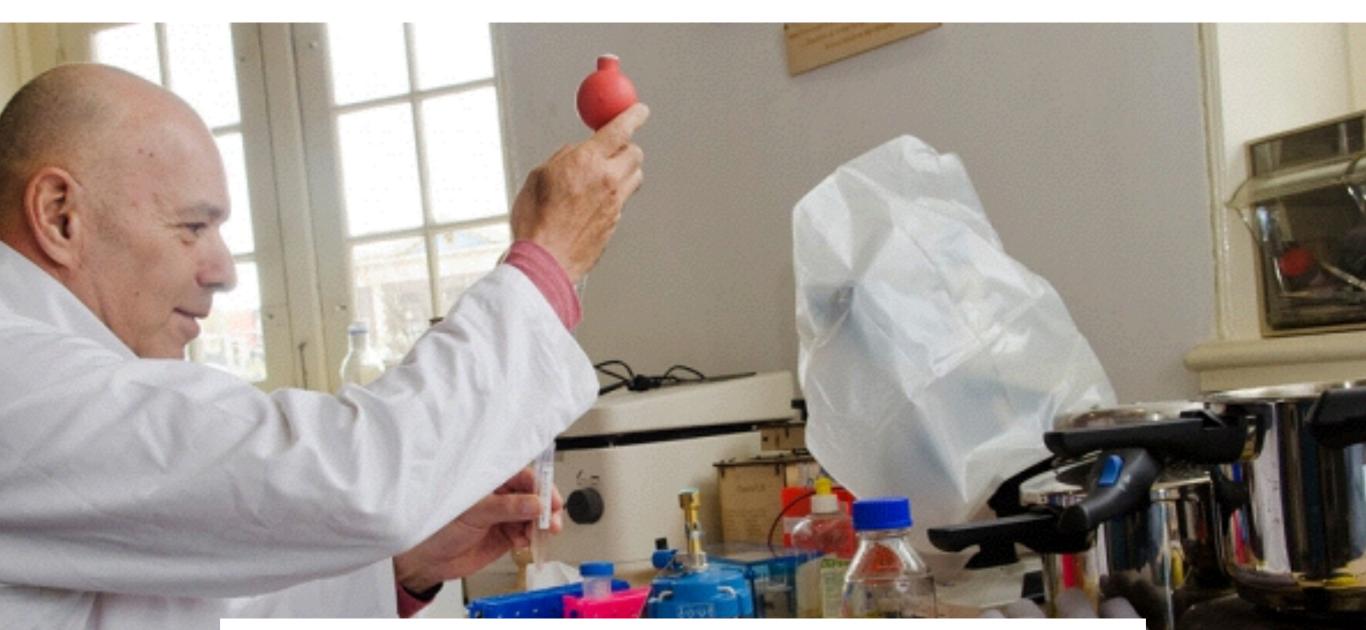


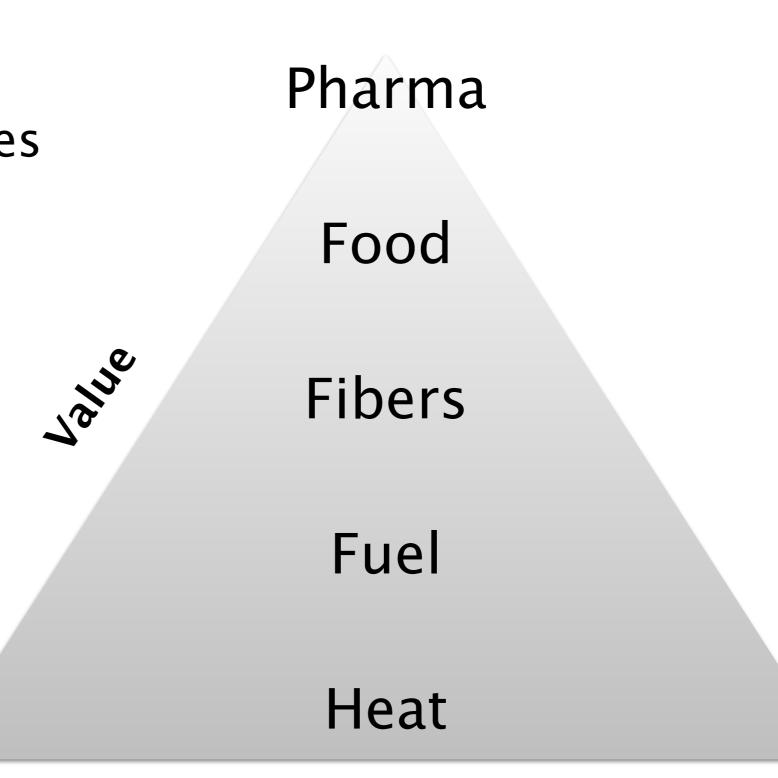
institute for art, science and technology



BioHack Academy Bioreactor Design

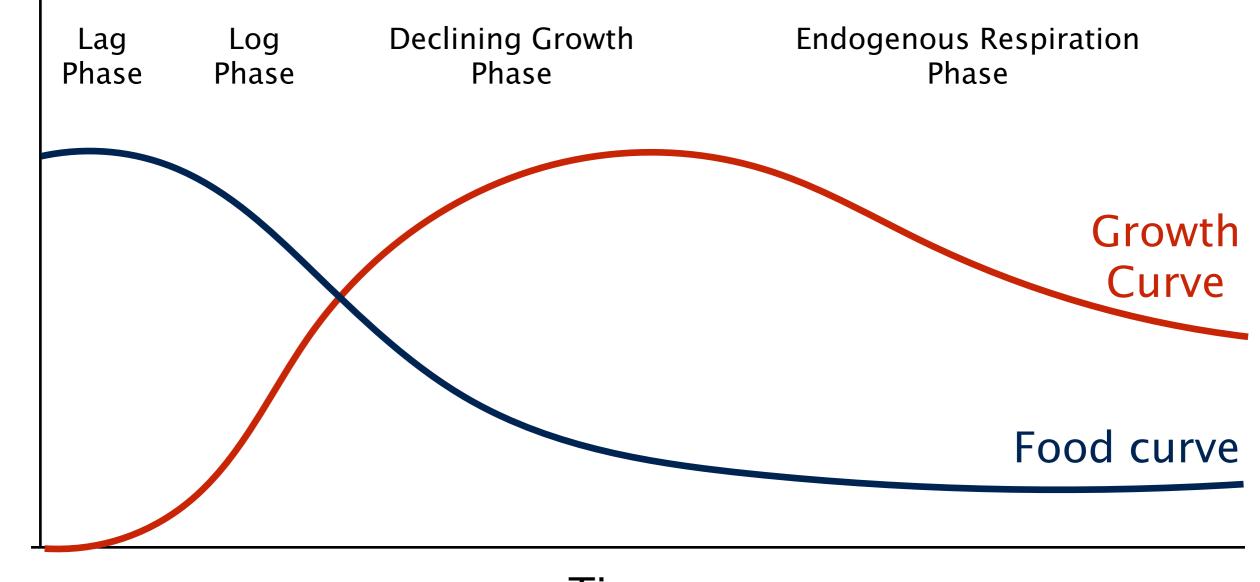


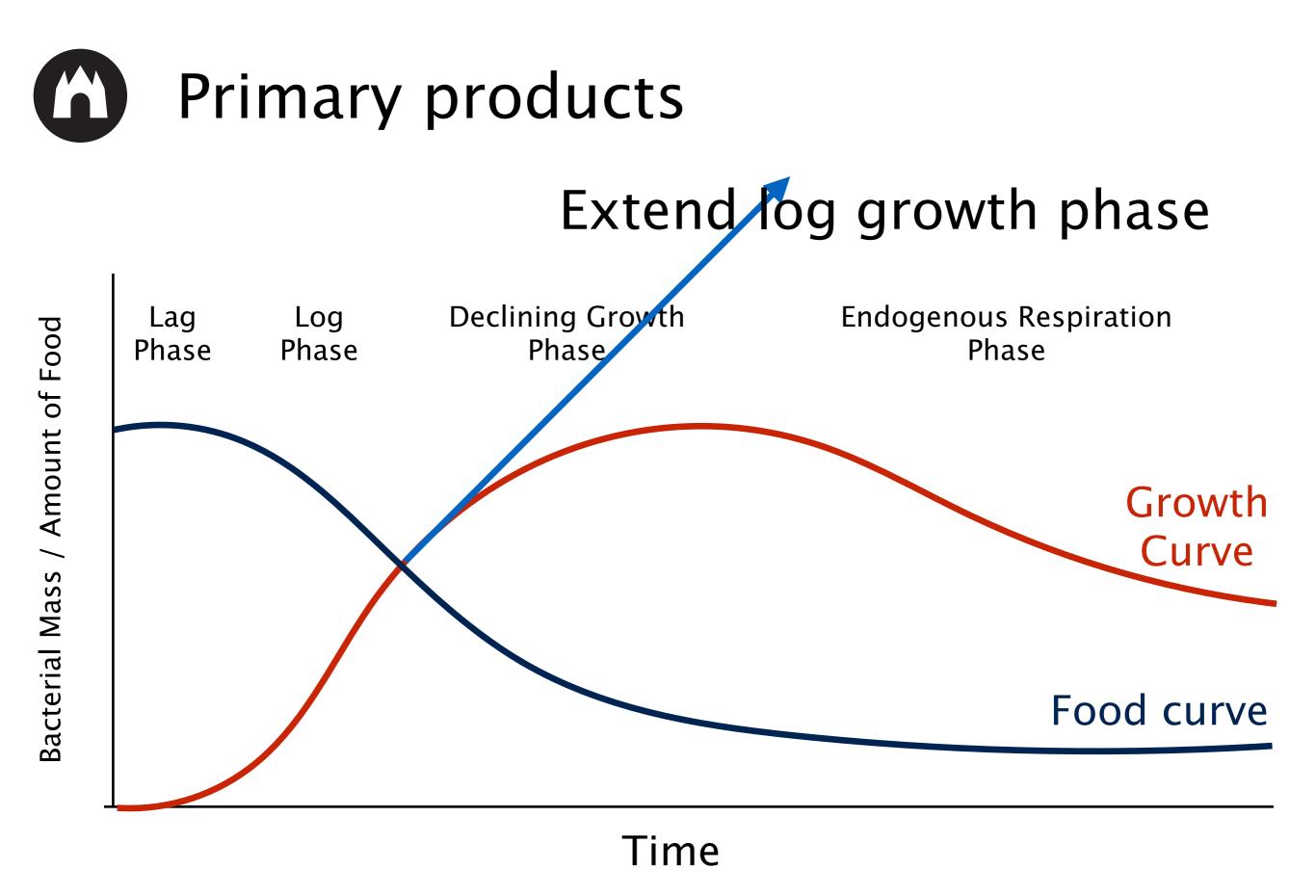
- Antibiotics
- Steroids / hormones
- Vitamins
- Proteins
- Sugars
- Acids



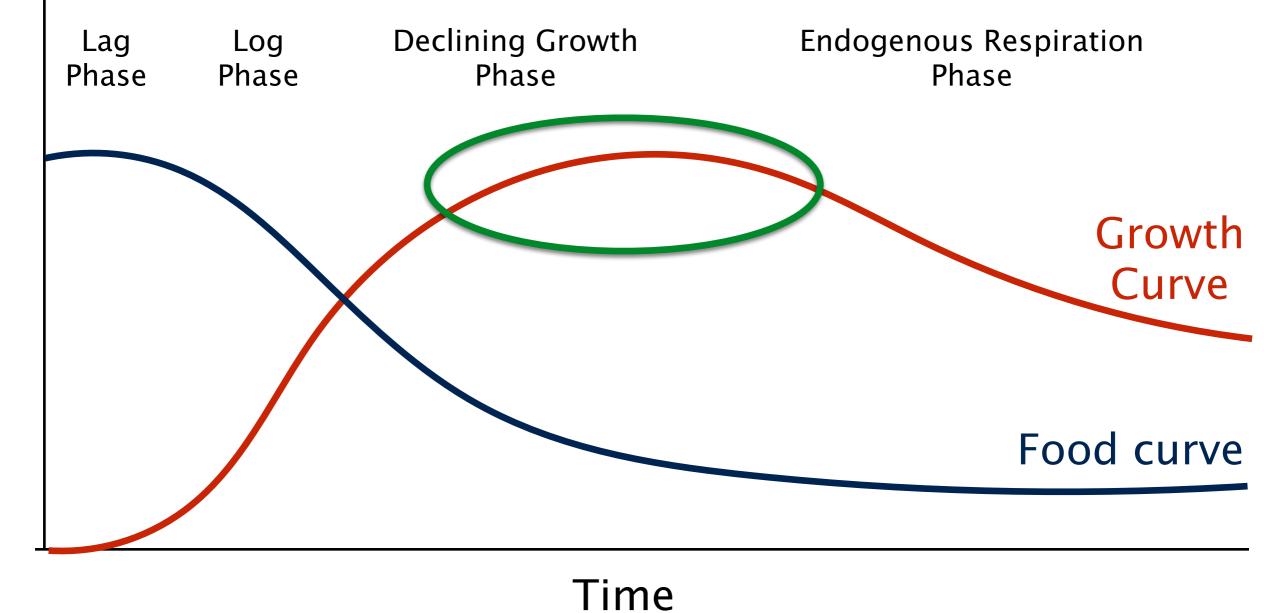
Volume



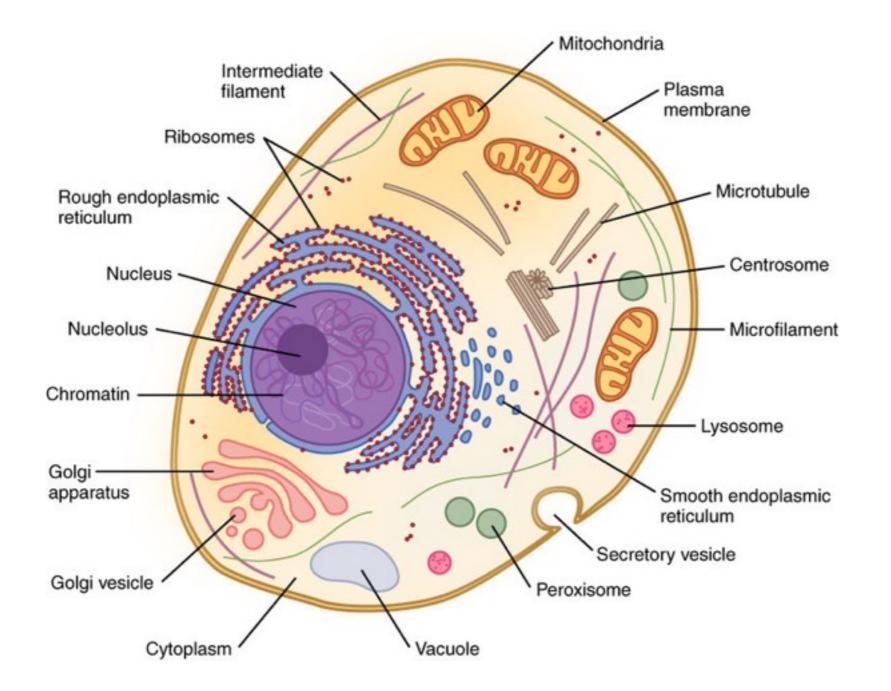




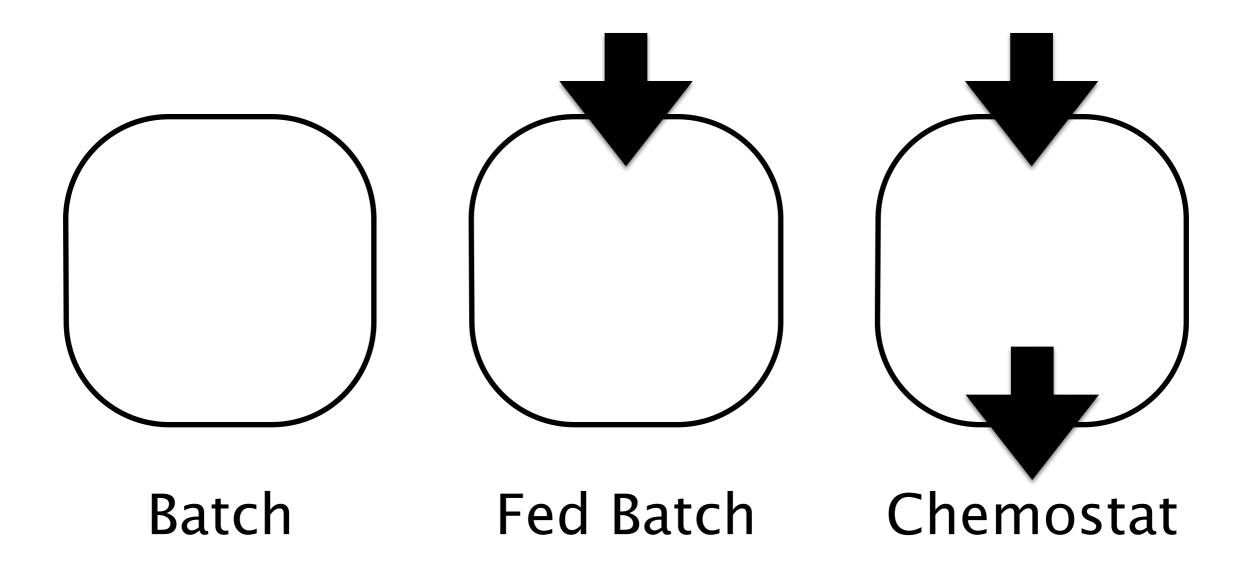




Intra vs extra cellular





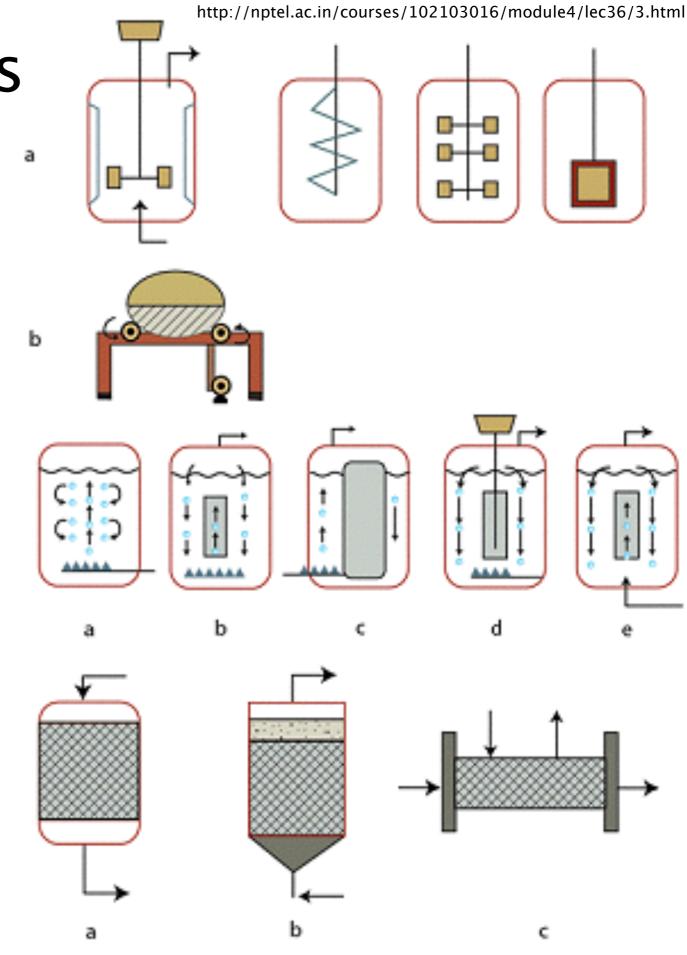




- Simple design
- Many cells per volume
- Uniform distribution
- Simple oxygen supply
- Low energy use



- Stirrer tank
- Air-lift
- Membrane
- Immobilized cells
- Cell culture
- Solid state
- Photobioreactor
- Microbioreactors
- Animals

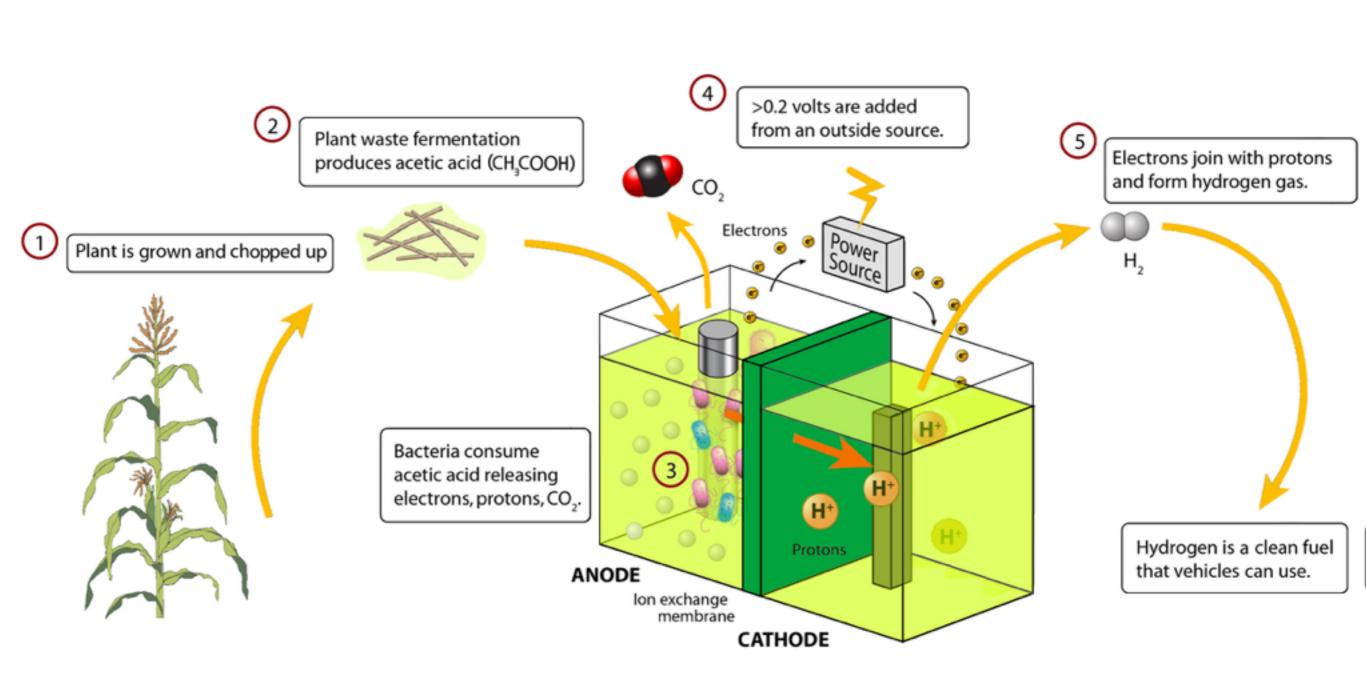




http://www.instructables.com/id/An-Algae-Bioreactor-from-Recycled-Water-Bottles/

http://www.instructables.com/id/Biogas-at-home-Cheap-and-Easy/





Membrane reactor: Fuel Cells

MICROBIAL ELECTROLYSIS CELL

PublicDomain



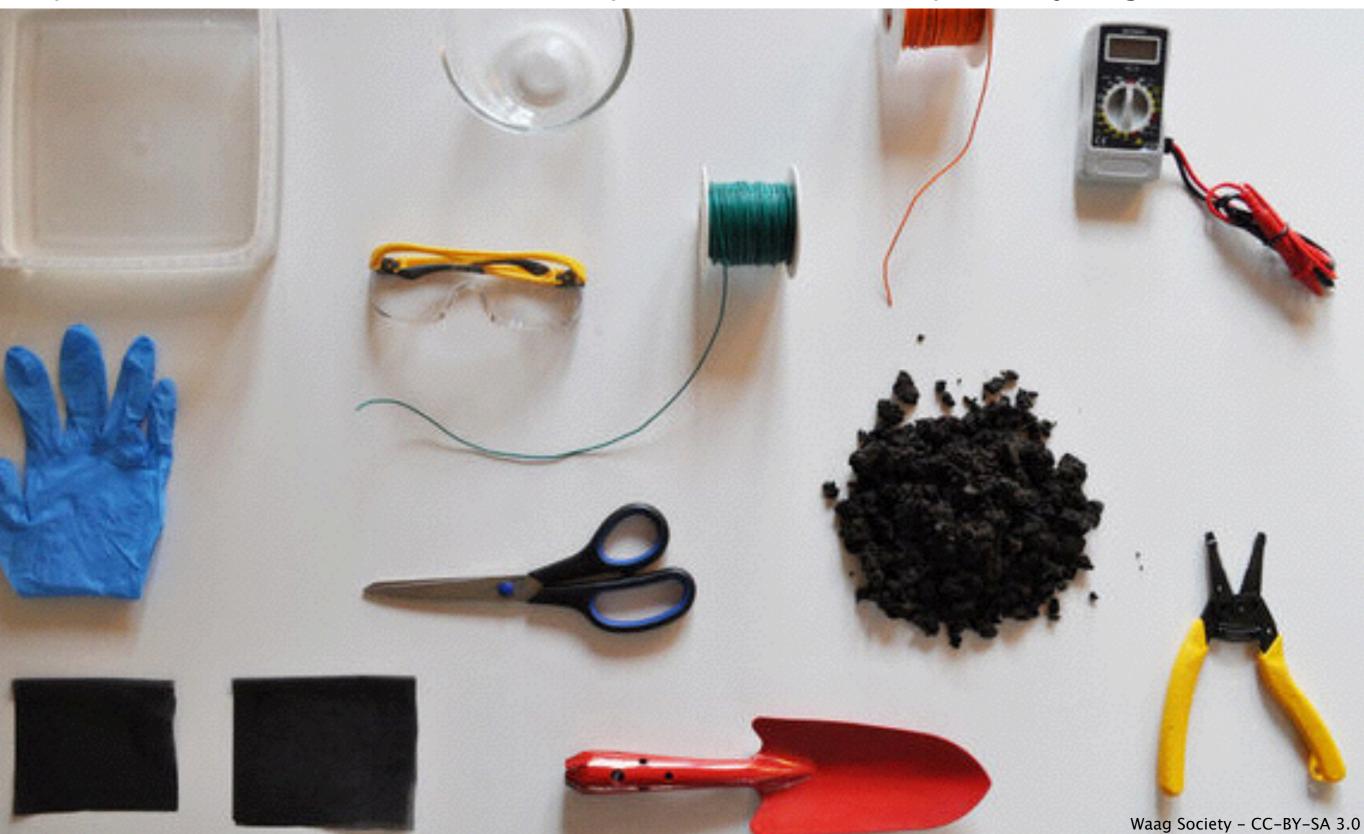


10, CATHODE H₂O (Nutrients) CH3COOH ANODE -ANODE e BIOFILM

MFCGuy2010 - CC-BY-SA 3.0

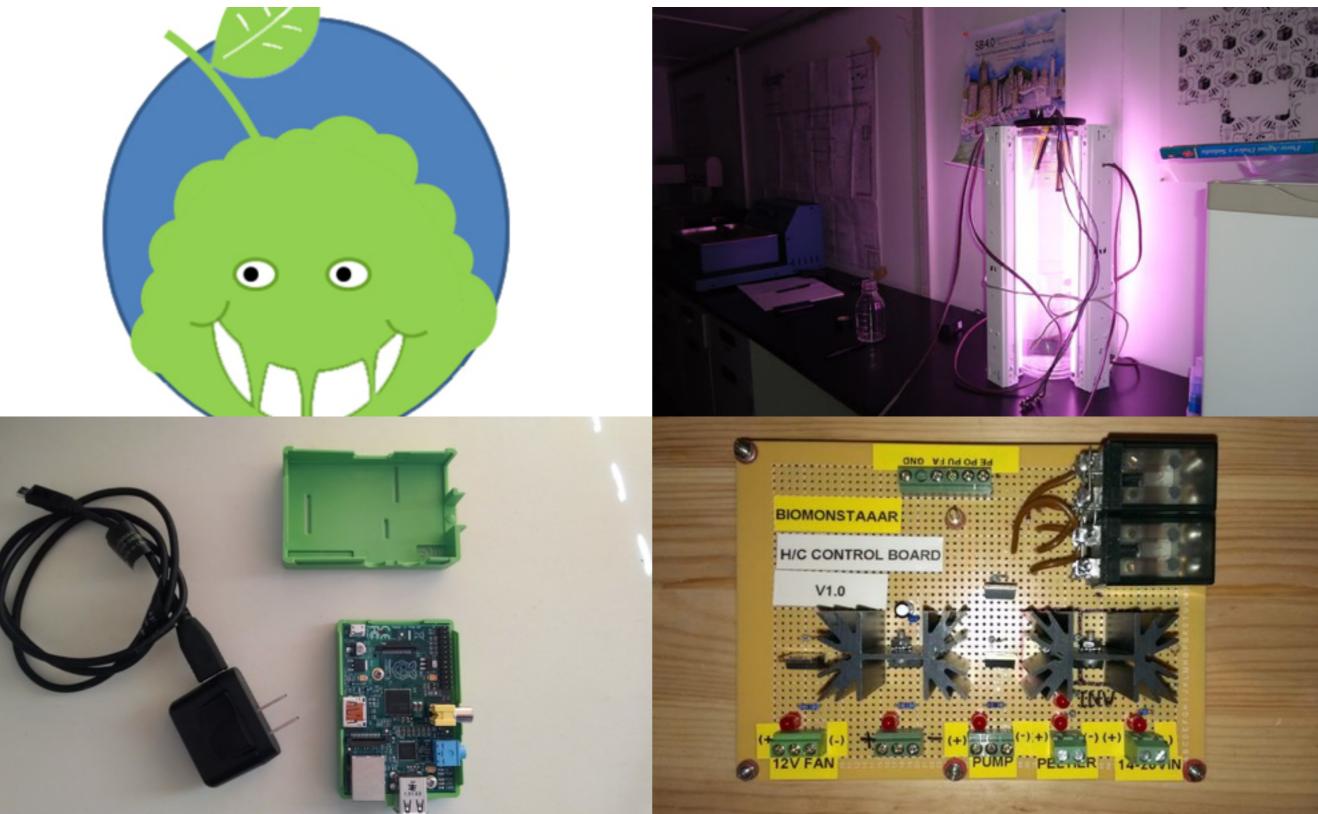


http://www.instructables.com/id/Create-your-own-mud-battery-Bouw-je-eigen-modderba/

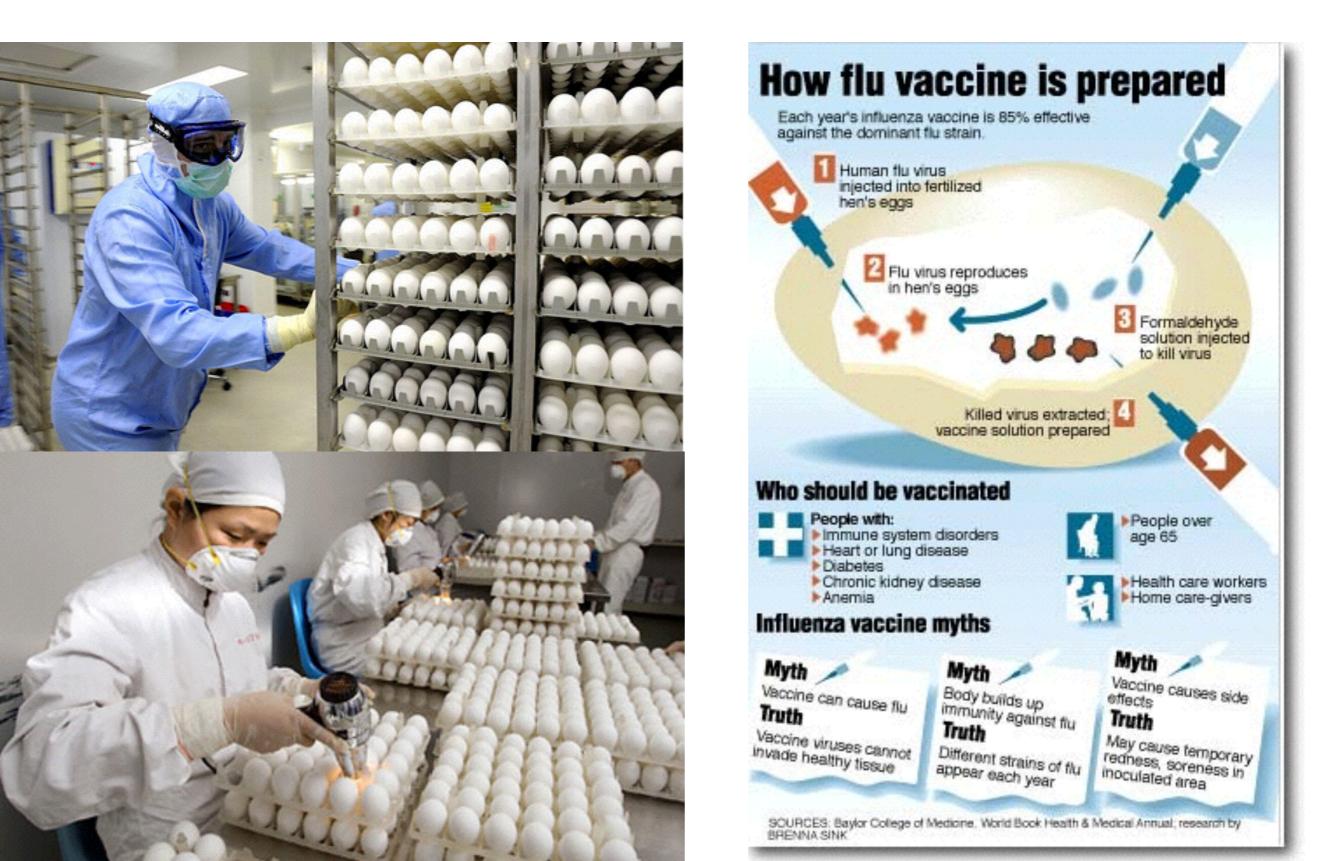




http://biomonstaaar.com



G Flu vaccine production in eggs





Cervarix Active Ingredient

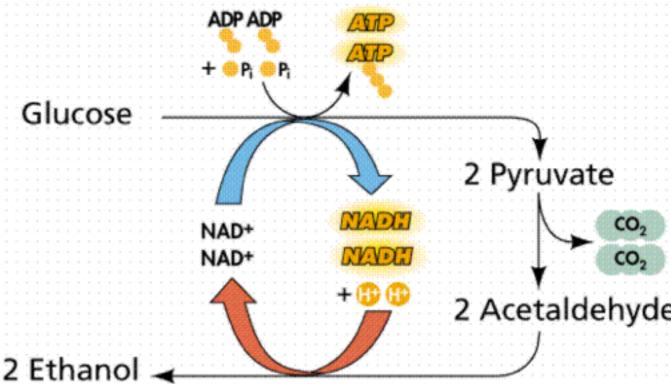
 Cervarix, HPV-16/18 L1 AS04 vaccine contains recombinant C-terminally truncated major capsid L1 proteins of HPV types 16 and 18 as active ingredients

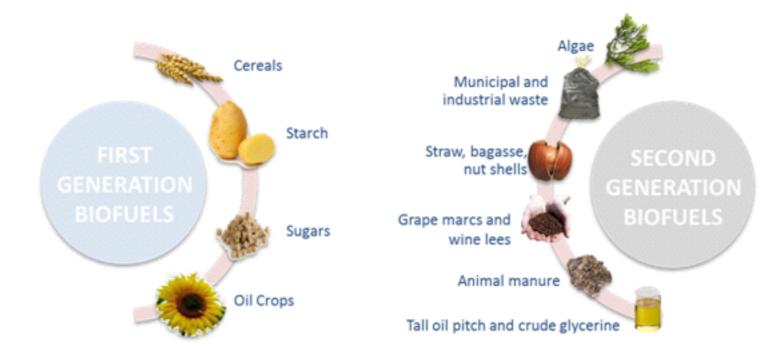
The first vaccines for humans from Baculovirus
Expression System

1 pentamer

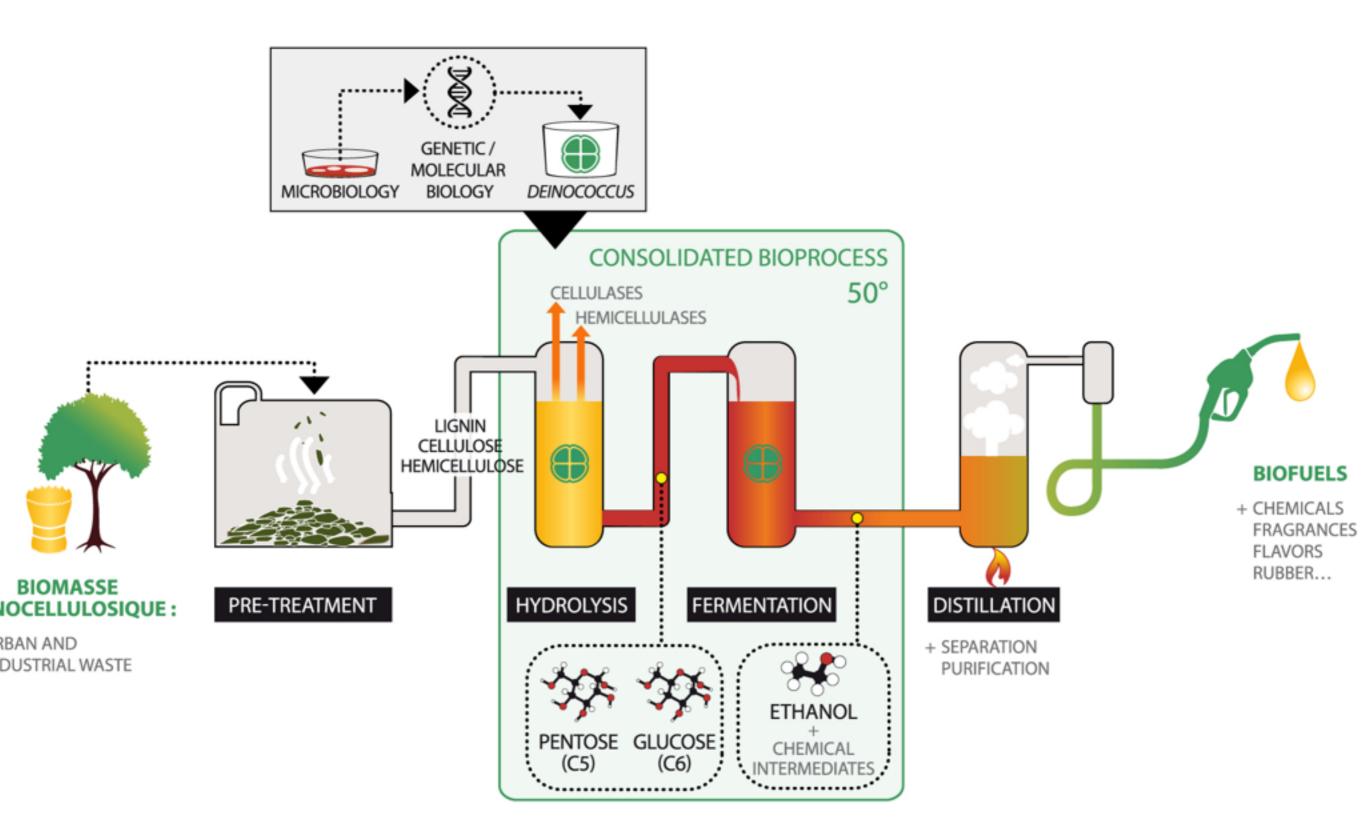




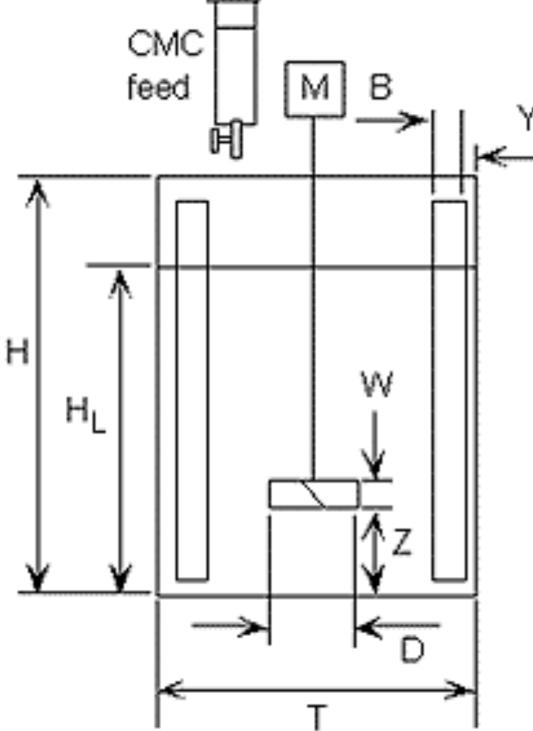








Geometry of standard stirred tank for aerobic reactions



Reactor Configuration		
Tank diameter	Т	105 mm
Baffles		4 number
Baffle width	в	T/12
Baffle spacing	Y	T/60
Impeller diameter	D	Т/З
Bottom clearance	Z	Т/З
Liquid depth	հլ	Т
Number of blades	n	4
Blade width	w	D/5
Blade angle	α	45 °

Volume = 800 mL

Brazilian Journal of Chemical Engineering



Example design

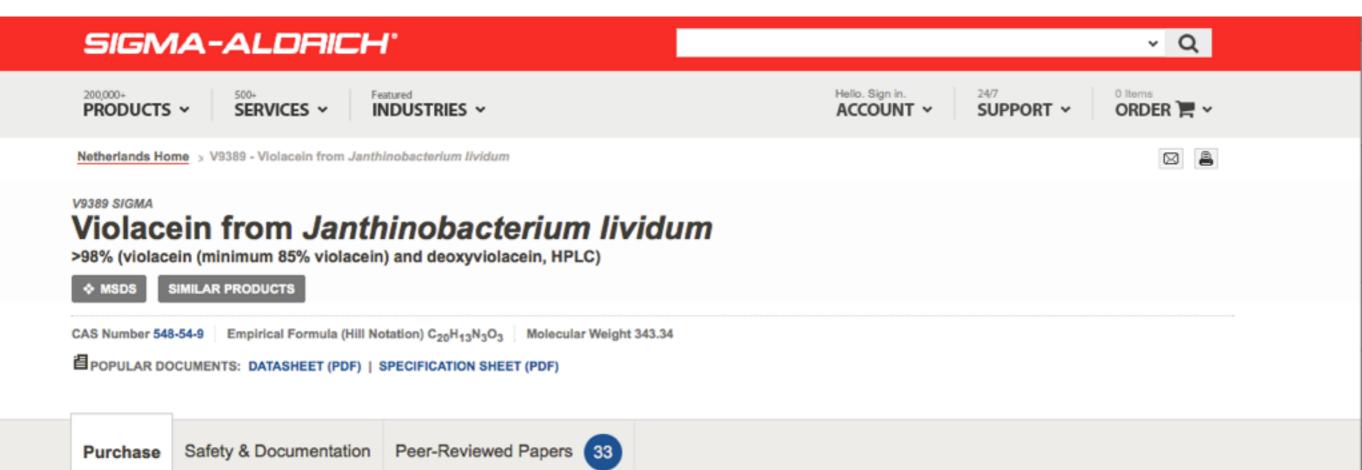
Violacein production

My search for J. lividum

- "Janthinobacterium lividum" +
 - "growth conditions"
 - "violacein pathway"
 - "violacein genes"
 - "patent"
 - "yield"
 - "inhibition"
 - "extraction"



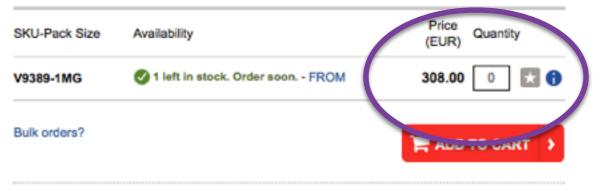




Properties

Apoptosis Inducers, Apoptosis and Cell Cycle, Bioactive Small Molecule Alphabetical Index, Bioactive Small Molecules, Cell Biology, More
>98% (violacein (minimum 85% violacein) and deoxyviolacein, HPLC)
H ₂ O: insoluble
acetone: soluble
ethanol: soluble

Price and Availability



Protein-Protein Interaction Webinar Series

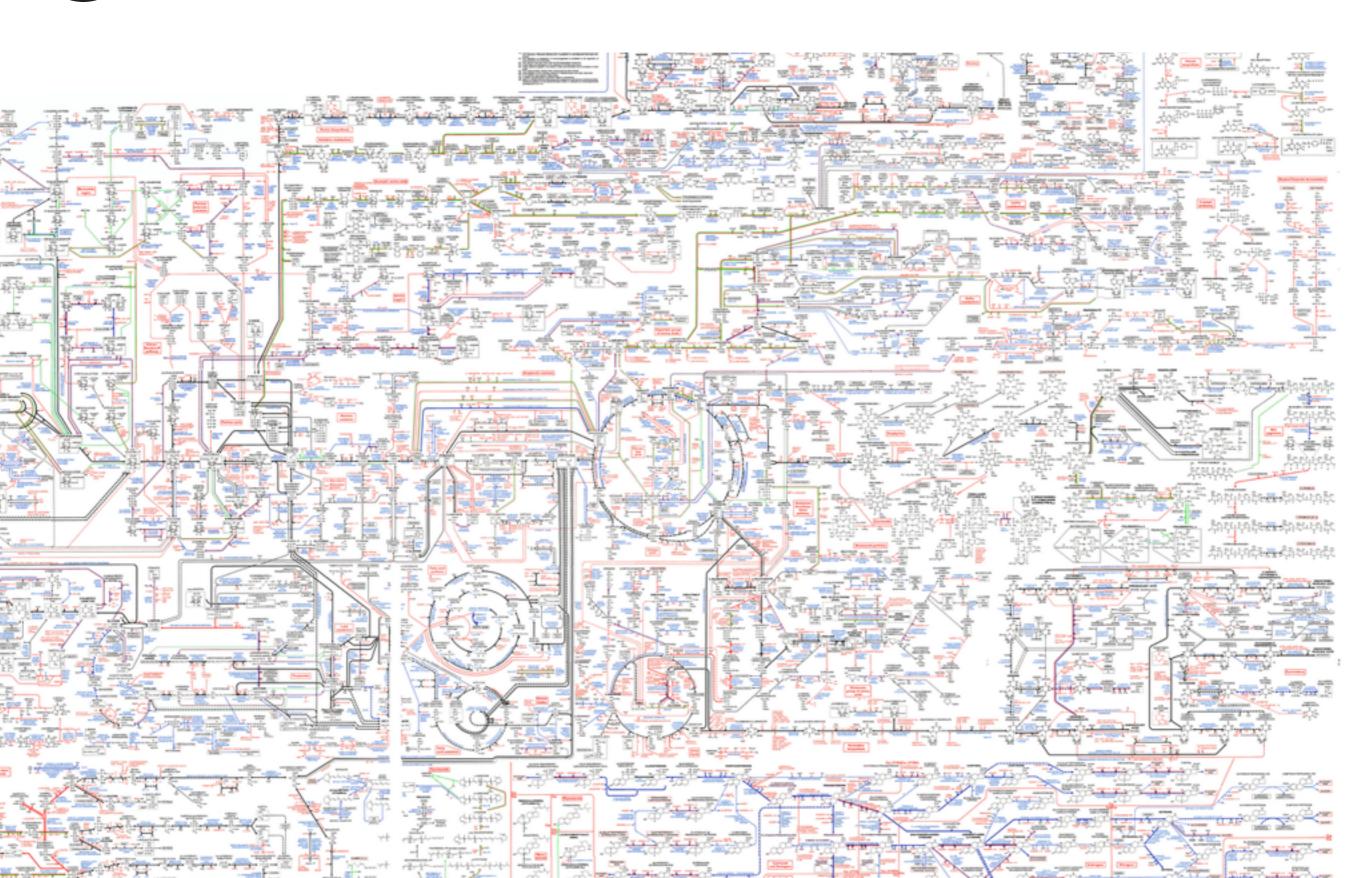
G Janthinobacterium lividum

Wikipedia tells me:

- Gram negative
- Aerobic

Violacein production from glycerol





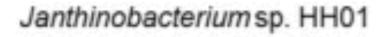


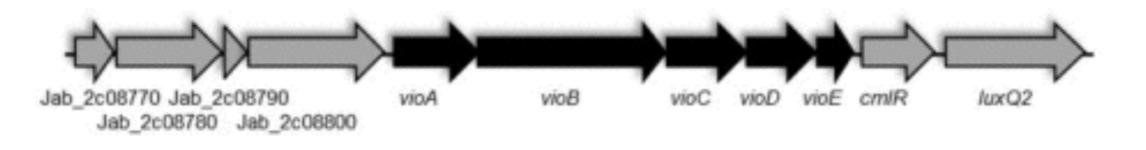
$C_{3}H_{5}O_{3}^{-} + 3O_{2} + H^{+} -> 3CO_{2} + 3H_{2}O_{2}$

Acid is consumed

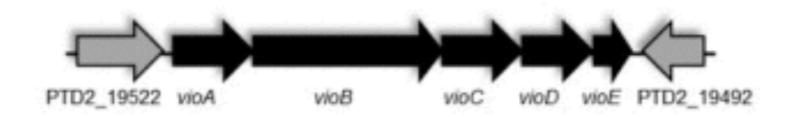


Hornung et al. - The Janthinobacterium sp. HH01 Genome Encodes a Homologue of the V. cholerae CqsA and L. pneumophila LqsA Autoinducer Synthases (2013)

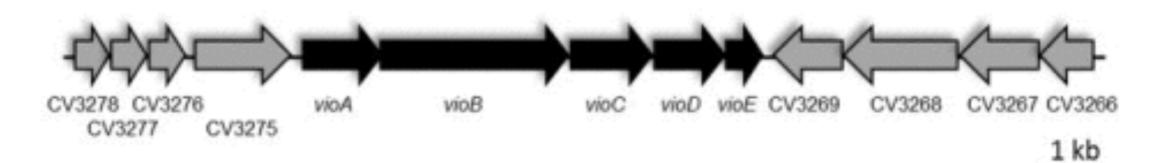




Pseudoalteromonas tunicata D2

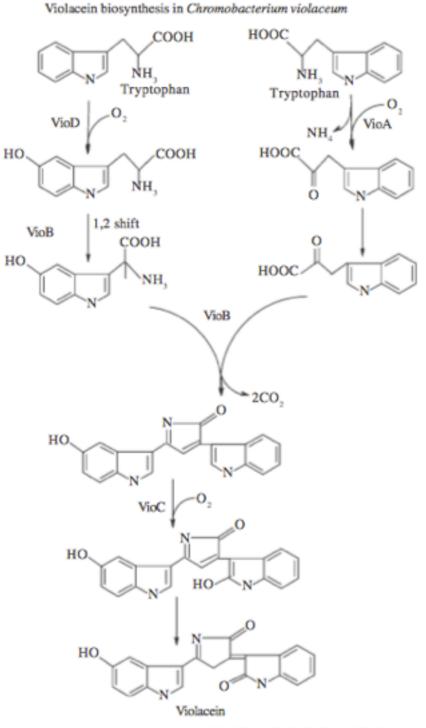


Chromobacterium violaceum ATCC 12472



Production pathway?

Tryptophan



89

Figure 2. Violacein biosynthesis, as proposed by August et al., 2000. VioA, VioB, VioC, and VioD are the gene products of the biosynthesis operon, encoding nucleotide-dependent monooxygenases and a protein similar to a polyketide synthase (VioB).

Antonio, R.V. and Creczynski-Pasa, T.B. (2004) Genetics analysis of violacein biosynthesis by C. violaceum. Genet Mol Res 3, 85-91.



• *J. lividum* produces a metallo- β -lactamase conferring resistance to

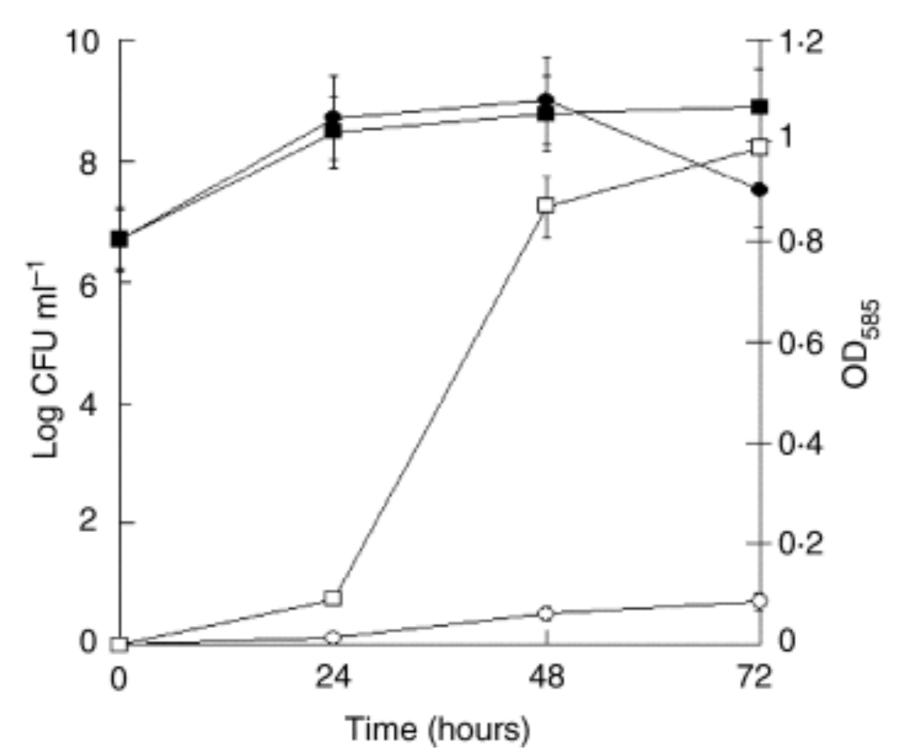
several β -lactam antibiotics

Rossolini, G.M., Condemi, M.A., Pantanella, F., Docquier, J.D., Amicosante, G. and Thaller, M.C. (2001) Metallo-β-lactamase producers in environmental microbiota: new molecular class B enzyme in Janthinobacterium lividum. Antimicrob Agents Chemother 45, 837-844.

- Violacein:
 - C_{20} - H_{13} - N_3 - O_3
 - molecular weight of 343.33
 - insoluble in water
 - soluble in alcohols as methanol, ethanol and acetone
 - maximal absorption in a solution of methanol is at 585 nm

Blosser, R.S. and Gray, K.M. (2000) Extraction of violacein from Chromobacterium violaceum provides a new quantitative bioassey for N-acyl homoserine lactone autoinducers. J Microbiol Methods 40, 47-55.





Pantanella, F., Berlutti, F., Passariello, C., Sarli, S., Morea, C. and Schippa, S. (2007), Violacein and biofilm production in *Janthinobacterium lividum*. Journal of Applied Microbiology, 102: 992–999. doi: 10.1111/j.1365-2672.2006.03155.x

Production conditions

Growing the bacteria in culture took 5 days before the culture would turn purple due to *J. lividum* forming a biofilm in the media. Large culture growth by embedding sterile cotton mats in sterile 2L bottles with nutrient media with the added glycerol and L-tryptophan **(fig. 2)** that showed purple coloring after 48 hour incubation [9]. The mats were extracted after 5 days to harvest the violacein. Yield of violacein from after crude methanol extraction and low was about 10mg.



Figure 2: Violacein optimization. 1% Glycerol and 250µM L-tryptophan were added to the nutrient broth media to enhance pigment development. Cotton mats were used to allow bacteria to become sessile and produce violacein faster than with liquid cultures.



Process for the production of violacein and its derivative deoxyviolacein containing bioactive pigment from Chromobacterium sp. (MTCC5522)

EXAMPLE 1

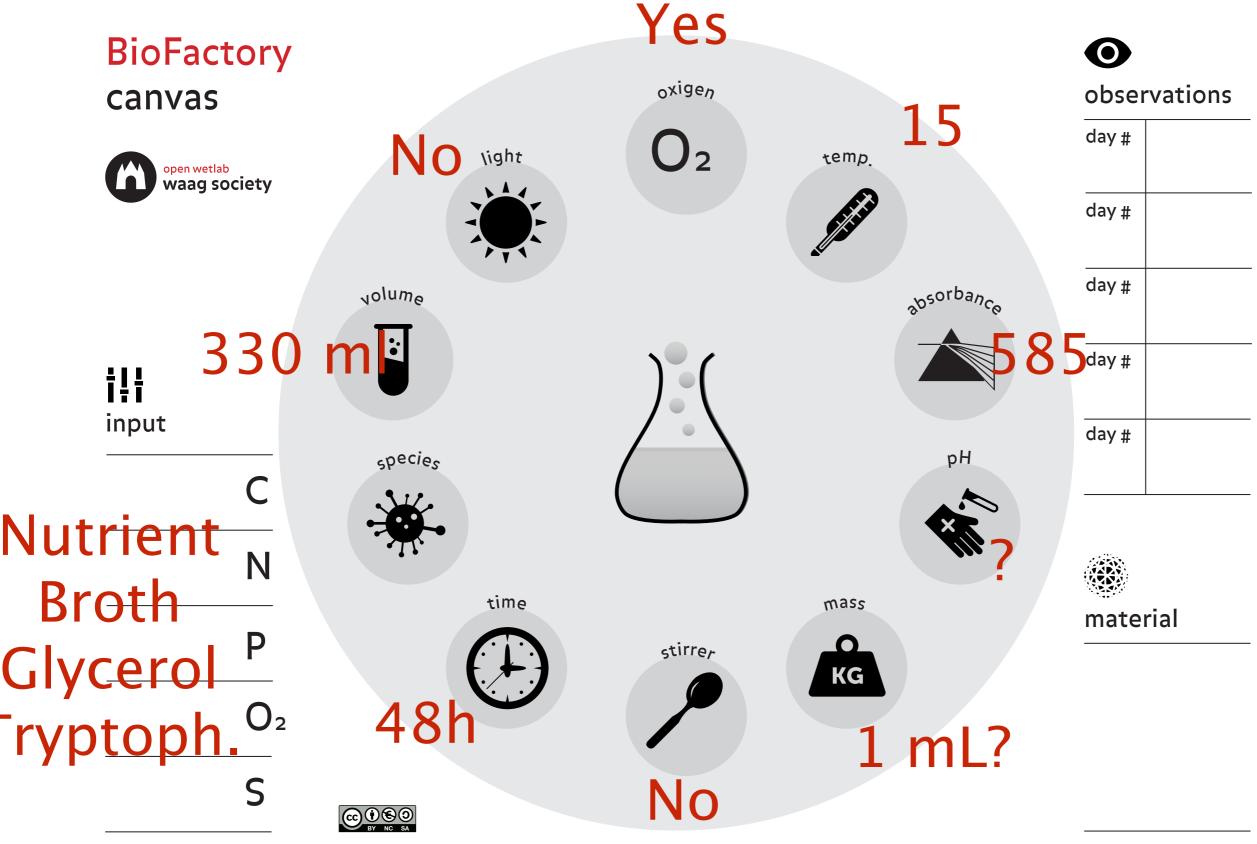
PRODUCTION AND EXTRACTION OF THE BIOACTIVE PIGMENT FROM THE CULTURE OF CHROMOBACTERIUM SP. NIIST-CKK-01

A loopful of 24 hrs old pure culture Chromobacterium sp. NIIST-CKK-01 from solid agar medium (LB agar or Nutrient agar) was inoculated with 50 ml of the growth medium (0.5% Yeast extract and 1.5% Peptone) taken in a 250 ml Erlenmeyer flask. Alternatively, 10% (v/v) of 24 hour old pure culture of Chromobacterium sp. NIIST-CKK-01 in LB broth was also used as inoculum. The pH of the medium was 7. The flasks inoculated with Chromobacterium sp. NIIST-CKK-01 were subsequently incubated in a rotary shaker at ambient temperature (30 °C) and 200 rpm for 24 hours. The deep purple purple-blue pigment starts appearing in the medium by about 6 hours of incubation and continued beyond biomass increase (Fig 1).

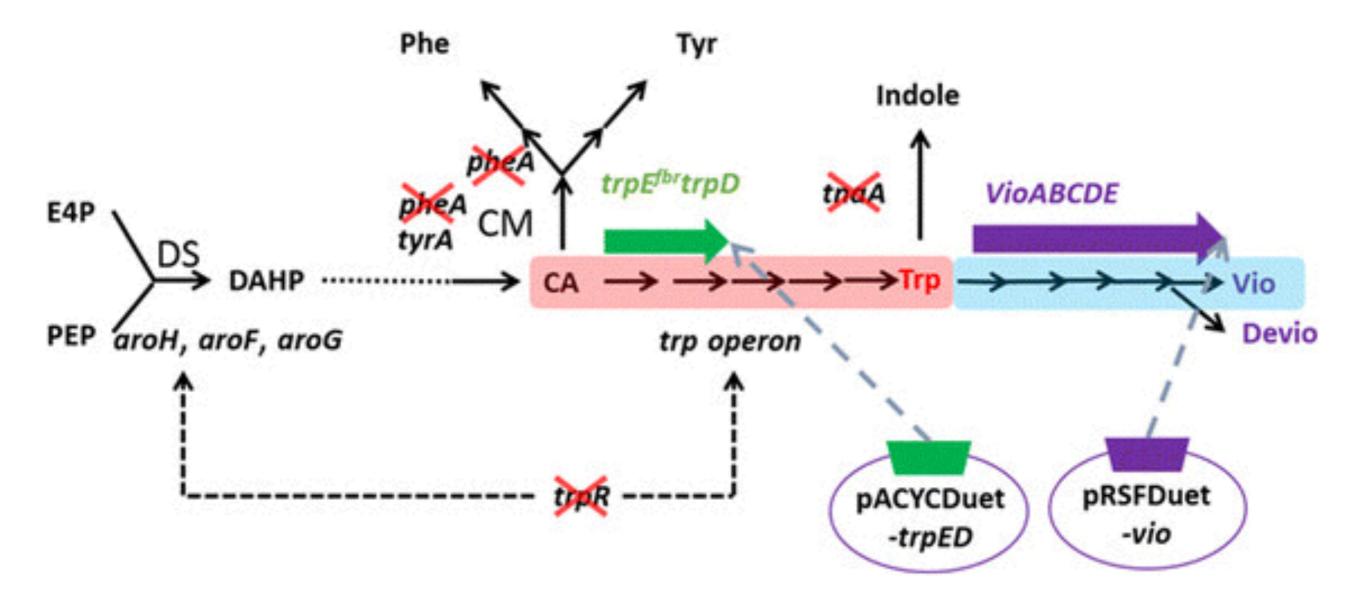
After 24 hrs of incubation, the bacterial biomass with pigment was centrifuged at 9676.8 x g and 4 °C for 10 minutes. After centrifugation, the clear supernatant was removed. The pellet containing biomass and pigment was mixed thoroughly with 5 ml of extra pure methanol. The mixture was centrifuged again at 9676.8 χ g and 4 °C for 10 minutes to separate the cell pellet from the solvent-pigment mixture. The pigment extraction was repeated twice using fresh solvent as described. All the pigment extracted solvent pooled together and the pigment was concentrated by normal vacuum drying in a desiccator. The quantity of biomass and pigment produced could be accounted by measuring optical density at 600 nm and 575 nm respectively. The yield of pigment by this method was about 1.0 g pigment/g of dry biomass in 24 hrs.

HPLC analysis is carried out for checking the purity of the pigment produced using an ODS column (Lichrospher-100; Merck) with acetonitrile (40%) at Iml/min as mobile phase and using UV-VIS detector at 575 nm (Figure 2). UV-VIS absorption spectra indicated maximum absorption at 575 nm, typical of violacein and its derivatives (Figure 3). EXAMPLE 2

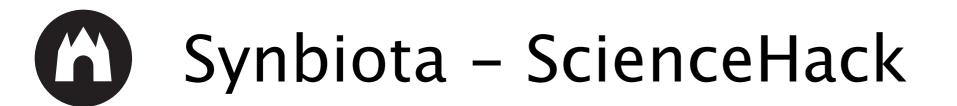








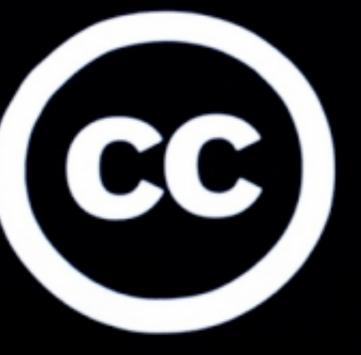
Fang et al. Microbial Cell Factories 2015 14:8 doi:10.1186/s12934-015-0192-x





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